## Relationship nitrification between soil Scottish Crop dynamics and population Research Institute Susan Mitchell, Jane Davidson, Ron Wheatley, Jim McNicol & Tim Daniell Plant-Soil Interface and <sup>1</sup>BioSS, Scottish Crop Research Institute, Invergowrie, Dundee DD2 5DA, UK. Introduction Previous work at SCRI showed potential nitrification rates in 3 neighbouring fields, 200m apart, to be significantly different, with distinct temporal patterns in each. Competitive PCR suggested nitrifying population sizes were different between, but did not change with time within, each field. There was no relationship between population size and PNR. Molecular analysis, using phylogenetic PCR, placed the nitrifiers in the Nitrosospira spp. cluster (Wheatley et al, EJSS,54 707-714). We have analysed spatial patterns of nitrification, denitrification and other soil characters in one field at SCRI. We present results of a preliminary molecular analysis of nitrifying populations (using an amo A gene fragment) from samples selected to give a range of activities. Method Potential nitrification was estimated and amo A gene fragments amplified, by PCR, from selected samples. Products were cloned and high-throughput sequencing, phylogenetics and multi-variant statistics applied to assess nitrifying population complexity. Potential nitrification is spatially and temporally variable. Inverse distance weighted plots represent the distribution of activity across the field site. Sampling locations for molecular analysis are indicated. A neighbor-joining tree (F84 model with gamma rates). Bootstrap analysis was performed (1000 repetitions) and Time Point shown where support 46 March 33 8.7 43 2.2 45 exceeds 69%. All 8.9 11.1 12.8 2.1 sequences detected 148 12 5 fall in Nitrosospira spp. group confirming 33 13.2 4.4 91 33 initial findings with 93 3.0 6.1 9.1 phylogenetic primers, the amo A fragment 148 12 utilised gave greater resolution. Summary of the distribution of sequence groups, expressed by percentage, and the total clone number. 27 single clone groups were found. Colour indicates sequence group 000 position on phylogenetic tree. Time point 1 Principal component analysis was performed to assess sequence group distribution. PCO 1 separates sample 148 from other samples (loadings suggest this is mainly due to changes in sequence froup 6 (18 clones) group 3, 5, 9, 10 and 13; note low sample size). PCO 2 separates respective time points for samples 93 and 96 but clusters time points of other samples (loadings Group 3 (69 clones) suggest this is mainly due to changes in sequence groups 4, 7, 12 and single clones). PCO 3 separates samples on the basis of time (loadings suggest this is mainly due to changes in sequence groups 1-4 and 14). **Summary** Group 2 (104 clones) Nitrification rates show temporal and spatial variation. Sequence analysis confirms no detectable presence of Nitrosomanas spp. within field. Preliminary analysis suggests variation in nitrifier population across both space and time.